

The effect of rhBMP-2 bonegraft on infrabony defects

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Abstract

rhBMP-2 bone graft has been used in orofacial defect and alveolar ridge augmentation. This clinical trial aims to evaluate the effect of rhBMP-2 compared with bioactive glass on the periodontal tissue regeneration in 2- and 3-wall infrabony periodontal defect

Method: 23 patients(male 13 and female 10) who had probing depth above 5 mm in one wall and the more of tooth walls received Biogran[®] bone grafts in 14 control sites and CowellBMP[®] in 13 experimental sites. The probing depth and gingival recession were measured at the base line, 3 month and 6 months after surgery.

Results: The experimental group showed the significant decrease in probing depth 3.78 mm and 3.96 mm on 3 and 6 months after surgery compared to 2.64 and 2.71 mm of the control group ($p < 0.05$). Gingival recession of both group were significantly decreased to 0.63 mm on 3 months and 0.65 mm on 6 months in experimental group after surgery and 0.50 mm on 3 months and 6 months in control group($p < 0.05$) but there were no difference between two groups. Probing attachment level were significantly increased to 2.93 mm in experiment group on 6 months compared to 4.04 mm in control group ($p < 0.05$).

Conclusion: Biogran[®] and CowellBMP[®] were effective in treatment of infra-bony periodontal defects. CowellBMP[®] was more significantly effective in decrease of probing depth and the increase of probing attachment level than Biogran[®].

The periodontal treatment aims to the decrease of periodontal pocket for easy plaque control and the regeneration of periodontal tissue in the defects. Gottlow and Nyman et al. used the barrier membrane with the various bone materials of autogenous, allogeneous, xerogenous and synthetic bone graft^{1,2}.

The autogenous bone graft is the gold standard, but the additional surgery and adverse effects limits the graft uptake practice³⁻⁶. The allogeneous bone graft has osteoinductive property, but the safety could not be

proven⁷. The xerogenous bone graft and synthetic bone graft have the osteoconductive property but do not have the osteoinductivity. Piorellini et al. reported that these grafts could not be successful in graft surgery.

The mesenchymal stem cells and growth factors have been recently tried in the periodontal tissue regeneration. The effect of recombinant human bone morphogenetic protein (rhBMPs) of growth factors on bone regeneration was reported by Urist in 1965⁸.

The DNA sequence and recombinant technique of

rhBMPs was identified and established⁹⁻¹¹. It was reported that the bone matrix saturated with rhBMP-2 induced the new isotopic bone in the muscle of mice according to the transformed osteoblast from myoblasts^{12,21}.

rhBMPs, the superfamily of TGF-beta (transforming growth factor-beta), have the major key of mammalian skeleton development due to the effect on osteoblast, chondroblast and osteoclast^{13,14}. The bone regeneration effect of rhBMP-2 had been proven in the clinical trials¹⁶⁻²⁰. It was reported the effect of rhBMP-2 on the periodontal tissue regeneration was positive in the animal experience²²⁻²⁶.

The effect of rhBMP-2 on the periodontal tissue regeneration in human was not studied. This clinical trial aims to evaluate the effect of rhBMP-2 compared with allograft on the periodontal tissue regeneration in 2- and 3-wall infrabony periodontal defect

I . Material and methods

1. Study Subject

The study was designed as a prospective, randomized controlled study documenting the response of the rhBMP-2 bone graft material and bioactive glass bone graft material in the periodontal department of Dankook dental school, Cheonan, Korea.

23 patients(male 13 and female 10) who had probing depth above 5 mm in one wall and the more of tooth which have over 2 mm keratinized gingiva and under 1 degree mobility received Biogran[®] bone grafts in 14 control sites and CowellBMP[®] in 13 experimental sites.

2. Material

The control group was treated with the bioactive glass bone graft material, Biogran[®](Biomet 3i Co., Ltd. Warsaw, United States) and the experiment group was done with the rhBMP-2 and HA/beta-TCP particles, CowellBMP[®](Cowellmedi Co., Ltd, Busan, Korea).

3. Treatment

1) Pre-operative treatment

The scaling, root planning and tooth brushing instruction were done at first visit. In case of traumatic from occlusion, the occlusal adjustment was done. At second visit, the periodontal control was evaluated and the additional oral hygiene instruction was done.

2) Surgical procedure

Infiltrate anesthesia was done in treatment site with 2% Lidocanine (1:80000 Epinephrine) injection. The sulcular incision and additional vertical incision (if need) was used. The full thickness flap was elevated with the periodontal elevator. The calculus removal and root planning were done in infrabony defect after the removal of granulation tissue. The planed root surface was treated with Tetracycline HCl for 2 minute. The defect was filled with Biogran[®] in control group and CowellBMP[®] in the experiment group. The flap was sutured with 4-0 Ethilon silk. The stitch were removed on 7~15 days after surgery(Fig 1).

After surgery, mouth rinsing with chlorhexidine 0.1% or 0.12%, twice daily for 10 days, was prescribed, together with the recommended medication prescribed by the surgeon (such as analgesics, anti-inflammatory compounds or antibiotics).

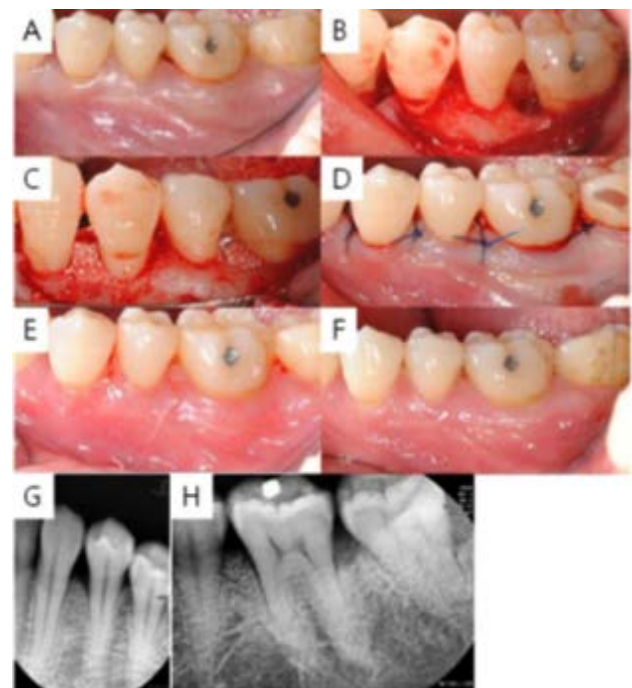


Figure 1. Surgical procedures and periapical view in experimental group

A : Preoperative status

B : Incision and flap reflection

C : Application of rhBMP-2

D : Suturing

E : Stitch out

F : Healing status after 3 months later

G : Periapical view of #34 teeth, 3 months after treatment

H : Periapical view of #36 teeth, 3 months after treatment

3) Measurement

All measurements were carried out by well-trained calibrated examiners independent from the surgeons placing the implants. The probing depth and gingival recession were measured on the pre- and post-operation, 3 months and 6 months after surgery at buccal, mesiobuccal, distobuccal, lingual, mesiolingual and distolingual sites with periodontal probe of 1 mm unit scale. Probing attachment level was measured on pre-operation, post-operation and 6 months after surgery.

4) Statistical method

The probing depth and gingival recession at the site of

maximal probing attachment level was analyzed on the pre-operative, post-operative, 3 months and 6 months after surgery with SPSS ver. 13.0. The change of measurement in two group according to periods was identified with Wicoxon signed ranks test. The difference between the measurement of two group according to periods was identified with Mann-Whitney test.

II. Result

1. The change of clinical index according to periods

1) The change between baseline and 3 months

The change of probing depth between baseline and 3 months was higher 3.78 ± 0.26 mm in experiment group than 2.64 ± 0.13 mm in control group with significant difference. ($p < 0.01$) (Table 1)

2) The change between 3 months and 6 months

The change of probing depth between 3 months and 6 months was 0.18 ± 0.09 mm in experiment group and 0.07 ± 0.07 mm in control group without significant difference. ($p > 0.05$)

Table 1. Changes of clinical indexes between baseline and 3 months (mm)

Parameter	Group	Baseline	3 Months	Difference	Significance
PD	control	6.58 ± 0.25	3.93 ± 0.22	2.64 ± 0.13	$< 0.001^*$
	experimental	7.62 ± 0.72	3.84 ± 0.32	3.78 ± 0.26	$< 0.001^*$
	<i>p</i> value			$< 0.01^*$	
GR	control	0.21 ± 0.11	0.71 ± 0.16	0.50 ± 0.14	$< 0.008^*$
	experimental	0.33 ± 0.25	0.96 ± 0.15	0.63 ± 0.21	$< 0.001^*$
	<i>p</i> value			NS	

PD : Probing pocket depth; GR : Gingival recession

NS : difference not statistically significant.

* = statistically significant difference ($P < 0.05$)

Table 2. Changes of clinical indexes between 3 months and 6 months (mm)

Parameter	Group	3 Months	6 Months	Difference	Significance
PD	control	3.93±0.22	3.86±0.21	0.07±0.07	0.32
	experimental	3.84±0.32	3.66±0.26	0.18±0.09	0.17
	<i>p</i> value			NS	
GR	control	0.71±0.16	0.71±0.16	0.00	1.00
	experimental	0.96±0.15	0.98±0.12	0.02±0.05	0.46
	<i>p</i> value			NS	

PD : Probing pocket depth; GR : Gingival recession

NS : difference not statistically significant.

* = statistically significant difference (P <0.05)

3) The change between baseline and 6 months

The change of probing depth between baseline and 6 months was higher 3.96±0.23 mm in experiment group than 2.71±0.13 mm in control group with significant difference.(p<0.05)

Probing attachment level were significantly increased to 2.93±0.02 mm in experiment group on 6 months compared to 4.04±0.13 mm in control group (p<0.05).(Table 2)

Table 3. Changes of clinical indexes between baseline and 6 months (mm)

Parameter	Group	Baseline	6 Months	Difference	Significance
PD	control	6.58±0.25	3.86±0.21	2.71±0.13	<0.001*
	experimental	7.62±0.72	3.66±0.26	3.96±0.23	<0.001*
	<i>p</i> value			<0.05*	
GR	control	0.21±0.11	0.71±0.16	0.50±0.14	<0.008*
	experimental	0.33±0.25	0.98±0.12	0.65±0.13	<0.005*
	<i>p</i> value			NS	
PAL	control	9.07±0.32	6.14±0.29	2.93±0.22	<0.001*
	experimental	9.92±0.22	5.88±0.21	4.04±0.13	<0.001*
	<i>p</i> value			<0.01*	

PD : Probing pocket depth; GR : Gingival recession;

PAL : Probing Attachment Level; NS : difference not statistically significant.

* = statistically significant difference (P <0.05)

2. The change of clinical index at each periods(Fig.2)

The change of probing depth between baseline and 3 months was higher 3.78 ± 0.23 mm in experiment group than 2.64 ± 0.13 mm in control group with significant difference.($p < 0.05$)

The change of probing depth between baseline and 6 months was higher 3.96 ± 0.23 mm in experiment group than 2.71 ± 0.13 mm in control group with significant difference.($p < 0.05$)

The change of gingival recession between baseline and 3

months was higher 0.63 ± 0.21 mm in experiment group than 0.5 ± 0.14 mm in control group without significant difference.

The change of gingival recession between baseline and 6 months was higher 0.65 ± 0.13 mm in experiment group than 0.5 ± 0.14 mm in control group without significant difference.

Probing attachment level were significantly increased to 2.93 ± 0.02 mm in experiment group on 6 months compared to 4.04 ± 0.13 mm in control group ($p < 0.05$).

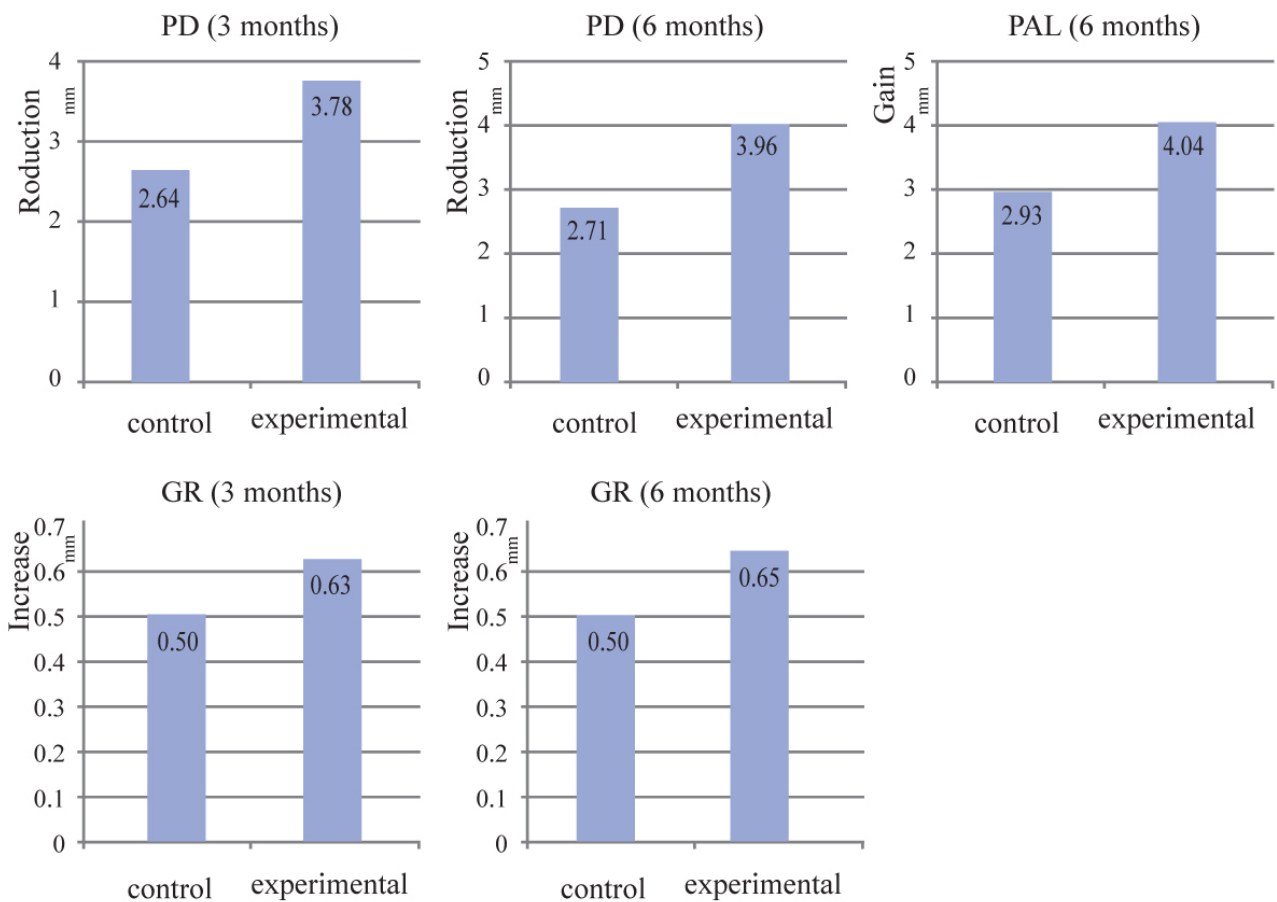


Fig 2. Changes of clinical indexes at each time from baseline
 PD : pocket depth, PAL : probing attached level, GR : Gingival recession

* Compared with control group ($p < 0.05$)

III. Discussion

rhBMP-2 bone graft was reported to be effective in oral maxillofacial bone defects, sinus bone augmentation and socket preservation²⁷⁻²⁹. It was reported that the longitudinal marginal bone change of implant placement and simultaneous rhBMP-2 bone graft was stable^{20,80,81}. Jung et al. (2008) reported that the rhBMP-2 experiment group had been more effective than the control group in the 43 of total 45 studies of periodontal tissue regeneration, socket preservation and peri-implant defect regeneration and the growth factor like as BMP-7, GDF-5 and PDGF⁸².

It was reported that rhBMP-2 had the potential property of the regeneration of cement and periodontal ligament²²⁻²⁶. It was reported that the cementoid tissue regenerated by rhBMP-2 prevented the epithelial migration⁸⁸. But Sigurdsson et al. reported that rhBMP-2 induced the ankylosis in periodontal regeneration site^{84,86}. It was reported that the high dose of rhBMP-2 could induce the dentinal resorption⁸⁸. The bone regeneration effect of rhBMP-2 was supported by various studies. In contrast, there are the controversy in periodontal tissue regeneration.

Bioactive glass, Biogran[®] used in the control group of this study is reported to bind bone and contact with soft tissue^{87,88}. Wilson and Low et al. reported that Bioactive glass has the osteoconductive property and effectiveness in infrabony periodontal defect⁸⁹. Zamet et al. reported that the probing depth was reduced to 3.82 mm and the gingival recess was 1.09 mm in Biogran[®] treatment group^{40,41}. Lovelance et al. reported that Biogran[®] and DFDBA reduced the probing depth without significant difference⁴². Therefore, Biogran[®] was used in this study.

The carrier of rhBMP-2 is composed of the absorbable collagen sponge, decalcified bone matrix and synthetic bone graft material. CowellBMP[®] carrier is 70% beta-TCP/30%HA which the bone regeneration effect was proven⁴⁶.

The change of probing depth and gingival recession

between pre-operative and post-operative periods was effective in both group. The control group received Biogran[®] was same result as Lovelance et al. study⁴². The change of probing depth between baseline and 6 months was higher 3.96±0.23 mm in experiment group than 2.71±0.13 mm in control group with significant difference.($p<0.05$) Probing attachment level were significantly increased to 2.93±0.02 mm in experiment group on 6 months compared to 4.04±0.13 mm in control group ($p<0.05$).(Table 2)

In recent clinical studies, the adverse effect of rhBMP-2 was reported to the facial swelling which was increased to proportional of the dose of rhBMP-2^{46,48}. In this study, there were no this adverse effect due to the low dose of rhBMP-2.

IV. Conclusion

Biogran[®] and CowellBMP[®] were effective in treatment of infra-bony periodontal defects. CowellBMP[®] was more significantly effective in decrease of probing depth and the increase of probing attachment level than Biogran[®]. rhBMP-2 might to be effective in periodontal tissue regeneration even in this limited enroll clinical study.

Reference

1. Nyman, S., Lindhe, J., Karring, T., Rylander, H. New attachment following surgical treatment of human periodontal disease. *J Clin Periodontol.* 1982; 9: 290-6.
2. Gottlow, J., Nyman, S., Lindhe, J., Karring, T., Wennström, J. New attachment formation in the human periodontium by guided tissue regeneration. Case reports. *J Clin Periodontol.* 1986; 13: 604-16.
3. Hiatt, W. H., Schallhorn, R. G., Aaronian, A. J. The induction of new bone and cementum formation. IV. Microscopic examination of the periodontium following human bone and marrow allograft, autograft and nongraft periodontal regenerative procedures. *J Periodontol.* 1978; 49: 495-512.
4. Tonetti, M. S., Pini, P. G., Williams, R. C., Cortellini,

- P. Periodontal regeneration of human infrabony defects. III. Diagnostic strategies to detect bone gain. *J Periodontol.* 1993; 64: 269-77.
5. Bowers, G. M., Chadroff, B., Carnevale, R., Mellonig, J., Corio, R., Emerson, J., Stevens, M., Romberg, E. Histologic evaluation of new attachment apparatus formation in humans. Part III. *JPeriodontol.* 1989; 60: 683-93.
6. Mellonig, J. T., Bowers, G. M., Cotton, W. R. Comparison of bone graft materials. Part II. New bone formation with autografts and allografts: a histological evaluation. *J Periodontol.* 1981; 52: 297-302.
7. Piorellini, M., Nevins, A. Localized Ridge Augmentation/Preservation. A systemic review. *Periodontol.* 2003; 8: 321-327.
8. Urist, M. R. Bone: formation by autoinduction. *Science.* 1965; 150: 893-899.
9. Wozney, J. M., Rosen, V., Celeste, A. J., Mitscock, L. M., Whitters, M. J., Kriz, R. W., Hewick, R. M., Wang, E. A. Novel regulators of bone formation: molecular clones and activities. *Science.* 1988; 242: 1528-1534.
10. Celeste, A. J., Iannazzi, J. A., Taylor, R. C., Hewick, R. M., Rosen, V., Wang, E. A. and Wozney, J. M. Identification of transforming growth factor b family members present in boneinductive protein purified from bovine bone. *Proceedings of the National Academy of Sciences of the United States of America.* 1990; 87: 9843-9847.
11. Özkaynak, E., Rueger, D. C., Drier, E. A., Corbett, C., Ridge, R. J., Sampath, T. K. and Oppermann, H. OP-1 cDNA encodes an osteogenic protein in the TGF- β family. *EMBO Journal.* 1990; 9: 2085-2093.
12. Wang, E. A., Rosen, V., D'Alessandro, J. S., Bauduy, M., Cordes, P., Harada, T., Israel, D. I., Hewick, R. M., Kerns, K. M., LaPan, P., Luxenburg, D. P., McQuaid, D., Moutsatsos, I. K., Nove, J., and Wozney, J. M. Recombinant human bone morphogenetic protein induces bone formation. *Proceedings of the National Academy of Sciences of the United States of America.* 1990; 87: 2220-2224.
13. Katagiri, T., Takahashi, N. Regulatory mechanisms of osteoblast and osteoclast differentiation. *Oral Dis.* 2002; 8: 147-159.
14. Canalis, E., Economides, A. N., Gaggero, E. Bone morphogenetic proteins, their antagonists, and the skeleton. *Endocr. Rev.* 2003; 24: 218-235.
15. Wikesjö, U. M., Lim, W. H., Thomson, R. C., Cook, A. D., Wozney, J. M., Hardwick, W. R. Periodontal repair in dogs: evaluation of a bioresorbable space-providing macroporous membrane with recombinant human bone morphogenetic protein-2. *Journal of Periodontology.* 2003; 74: 635-647.
16. Wikesjö, U. M., Xiropaidis, A. V., Thomson, R. C., Cook, A. D., Selvig, K. A. and Hardwick, W. R. Periodontal repair in dogs: rhBMP-2 significantly enhances bone formation under provisions for guided tissue regeneration. *Journal of Clinical Periodontology.* 2003b; 30: 705-714.
17. Yudell, R. M., Block, M. S. Bone gap healing in the dog using recombinant human bone morphogenetic protein-2. *Journal of Oral and Maxillofacial Surgery.* 2000; 58: 761-766.
18. Hunt, D. R., Jovanovic, S. A., Wikesjö, U. M., Wozney, J. M. and Bernard, G. W. Hyaluronan supports recombinant human bone morphogenetic protein-2 induced bone reconstruction of advanced alveolar ridge defects in dogs. A pilot study. *Journal of Periodontology.* 2001; 72: 651-658.
19. Sykaras, N., Triplett, R. G., Nunn, M. E., Iacopino, A. M. and Opperman, L. A. Effect of recombinant human bone morphogenetic protein-2 on bone regeneration and osseointegration of dental implants. *Clinical Oral Implants Research.* 2001; 12: 339-349.
20. Jovanovic, S. A., Hunt, D. R., Bernard, G. W., Spiekermann, H., Wozney, J. M. and Wikesjö, U. M. Long-term functional loading of dental implants in rhBMP-2 induced bone. A histologic study in the canine ridge augmentation model. *Clinical Oral Implants Research.* 2003; 14: 793-803.
21. Nakashima, K., Zhou, X., Kunkel, G., Zhang, Z.,

- Deng, J. M., Behringer, R. R. and Crombrughe, B. The novel zinc finger-containing transcription factor osterix is required for osteoblast differentiation and bone formation. *Cell*. 2002; 108: 17–29.
22. Oda, S., Kinoshita, A., Higuchi, T., Shizuya, T., Ishikawa, I. Ectopic bone formation by biphasic calcium phosphate (BCP) combined with recombinant human bone morphogenetic protein-2 (rhBMP-2). *J Med Dent Sci*. 1997; 44: 53-62.
23. Steinberg, B., Chiego, D. J. Jr., Huizinga, P. J., Wozney, J. M., Wikesjö, U. M. Effect of human bone morphogenetic protein 2 implant on tooth eruption in an experimental design. *J Craniofac Surg*. 1999; 10: 338-41.
24. Kuboki, Y., Sasaki, M., Saito, A., Takita, H., Kato, H. Regeneration of periodontal ligament and cementum by BMP-applied tissue engineering. *Eur J Oral Sci*. 1998; 106: 197-203.
25. Saito, E., Saito, A., Kawanami, M. Favorable healing following space creation in rhBMP-2-induced periodontal regeneration of horizontal circumferential defects in dogs with experimental periodontitis. *J Periodontol*. 2003; 74: 1808-15.
26. Kusumoto, K., Bessho, K., Fujimura, K., Akioka, J., Okubo, Y., Wang, Y., Iizuka, T. and Ogawa, Y. Osteoinduction by recombinant human bone morphogenetic protein-2 in muscles of non-human primates. *J. Int. Med. Res*. 2002; 30; 251–259.
27. Boyne, P. J., Salina, S., Nakamura, A., Audia, F., Shabahang, S. Bone regeneration using rhBMP-2 induction in hemimandibulectomy type defects of elderly sub-human primates. *Cell Tissue Bank*. 2006; 7: 1-10.
28. Jung, R. E., Glauser, R., Schärer, P., Hämmerle, C. H., Sailer, H. F., Weber, F. E. Effect of rhBMP-2 on guided bone regeneration in humans. *Clin Oral Implants Res*. 2003; 14: 556-68.
29. Nevins, M., Kirker-Head, C., Nevins, M., Wozney, J. A., Palmer, R., Graham, D. Bone formation in the goat maxillary sinus induced by absorbable collagen sponge implants impregnated with recombinant human bone morphogenetic protein-2. *Int J Periodontics Restorative Dent*. 1996; 16: 8-19.
30. Matin, K., Senpuku, H., Hanada, N., Ozawa, H., Ejiri, S. Bone regeneration by recombinant human bone morphogenetic protein-2 around immediate implants: a pilot study in rats. *Int J Oral Maxillofac Implants*. 2003; 18(2): 211-7.
31. Triplett, R. G., Andrews, J. A., Hallmon, W. W. Management of peri-implantitis. *Oral Maxillofac Surg Clin North Am*. 2003; 15: 129-38.
32. Jung, R. E., Thoma, D. S., Hammerle, C. H. Assessment of the potential of growth factors for localized alveolar ridge augmentation: a systematic review. *J Clin Periodontol*. 2008; 35: 255-81.
33. Miyaji, H., Sugaya, T., Ibe, K., Ishizuka, R., Tokunaga, K., Kawanami, M. A Root surface conditioning with bone morphogenetic protein-2 facilitates cementum-like tissue deposition in beagle dogs. *J Periodontal Res*. 2010; 45: 658-63.
34. Sigurdsson, T. J., Lee, M. B., Kubota, K., Turek, T. J., Wozney, J. M., Wikesjö, U. M. Periodontal repair in dogs: recombinant human bone morphogenetic protein-2 significantly enhances periodontal regeneration. *J Periodontol*. 1995; 66: 131-8.
35. Steinberg, B., Chiego, D. J. Jr., Huizinga, P. J., Wozney, J. M., Wikesjö, U. M. Effect of human bone morphogenetic protein 2 implant on tooth eruption in an experimental design. *J Craniofac Surg*. 1999; 10: 338-41.
36. Miyaji, H., Sugaya, T., Kato, K., Kawamura, N., Tsuji, H., Kawanami, M. Dentin resorption and cementum-like tissue formation by bone morphogenetic protein application. *J Periodontal Res*. 2006; 41: 311-5.
37. Hench, L. L., Splinter, R. J., Allen W. C., Greenlee, T.K. Bonding mechanisms at the interface of ceramic prosthetic materials. *J Biomed Mater Res*. 1972; 2: 117-141.
38. Wilson, J., Noletti, D. Bonding of soft tissue to bioglass. *Handbook of bioactive ceramics*. 1990.
39. Wilson, J., Low, S. B. Bioactive ceramics for periodontal treatment: comparative studies in the Patus monkey. *J Appl Biomater*. 1992 ; 3: 123-9.

40. Zomet, J. S., Darbar, U. R., Griffiths, G. S., Bulman, J. S., Brägger, U., Bürgin, W., Newman, H. N. Particulate bioglass as a grafting material in the treatment of periodontal intrabony defects. *J Clin Periodontol.* 1997; 24: 410-8.
41. Froum, S. J., Weinberg, M. A., Tarnow, D. Comparison of bioactive glass synthetic bone graft particles and open debridement in the treatment of human periodontal defects. A clinical study. *J Periodontol.* 1998; 69: 698-709.
42. Lovelace, T. B., Mellonig, J. T., Meffert, R. M., Jones, A. A., Nummikoski, P. V., Cochran, D. L. Clinical evaluation of bioactive glass in the treatment of periodontal osseous defects in humans. *J Periodontol.* 1998; 69: 1027-35.
43. Jung, R. E., Weber, F. E., Thoma, D. S., Ehrbar, M., Hammerle, C. H. F. BMP-2 enhances bone formation when delivered by a synthetic matrix containing HA/TCP. *Clinical Oral Implants Research.* 2008; 19: 188–195.
44. Shah, M. M., Smyth, M. D. and Woo, A. S. Adverse facial edema associated with off-label use of recombinant human bone morphogenetic protein-2 in cranial reconstruction for craniosynostosis. Case report. *J. Neurosurg. Pediatr.* 2008; 1: 255-257.
45. Latzman, J. M., Kong, L., Liu, C. and Samadani, U. Administration of Human Recombinant Bone Morphogenetic Protein-2 for Spine Fusion May Be Associated With Transient Postoperative Renal Insufficiency. *Spine.* 2010; 35: 231-7.
46. Boakye, M., Mummaneni, P. V., Garrett M., Rodts, G., and Haid, R. Anterior cervical discectomy and fusion involving a polyetheretherketone spacer and bone morphogenetic protein. *J Neurosurg Spine.* 2005; 2: 521–525.